**Online Appendix**: "When good balance goes bad: A discussion of common pitfalls when using entropy balancing"

## <u>Appendix: Comparing Entropy Balancing (EB), Propensity Score Matching (PSM), and</u> <u>Ordinary Least Squares (OLS) Regression</u>

# 1. Overview

The goal of this appendix is to use a simulated dataset with a handful of variables to illustrate how three common analysis tools weight observations to construct a counterfactual. In doing so, this appendix builds on illustrations by Hainmueller (2012) and McMullin and Schonberger (2020) designed to illustrate how entropy balancing assigns observational weights to eliminate covariate imbalance across a treatment and control sample of interest.

We begin by generating data containing a treatment indicator t, a single covariate X, and an outcome variable Y. Using this simulated dataset, we use three analysis approaches to identify a control sample used to estimate a treatment effect. These approaches are:

Analysis Approach	Data Preprocessing Approach	Estimation Approach
OLS	None (ordinary least squares)	Regression (equal weighted
		over full sample)
PSM and OLS	Estimate propensity scores and match	Regression (equal weighted
	treatment and control sample	over propensity score
	observations on propensity scores	matched samples)
	without replacement to produce a	
	matched sample	
Entropy Balancing	Entropy balancing to identify weights	Regression (equal weights
and Weighted OLS	for each observation in the control	for treated sample and
	sample such that the weighted control	entropy balancing weights
	sample and treated sample have similar	for control sample over full
	covariate distributions	sample)

These three approaches typify research designs accounting researchers commonly use. In the interest of brevity, we do not examine related matching approaches in the simulation (e.g., coarsened exact matching [CEM]), as Morgan and Winship (2015) document that PSM outperforms a number of other matching approaches.

We provide Stata code in blocks of yellow and Stata output and figures in blocks of blue.

# 2. Update Stata, Install Packages, and Create Data

We first update Stata and install the entropy balancing package.

```
*-----*
* Update and Install software
*-----*
update all
ssc install ebalance, replace all
ssc install psmatch2, replace all
```

We next create 150 observations (50 treated and 100 control, denoted by a treatment indicator t) with one covariate X, constructed as a uniform random variable over the interval (0, 10) for the control sample (with t = 0) and over the interval (5, 10) for the treatment sample (with t = 1). As a result of this definition of X over a smaller interval for the treatment sample, treatment and control samples will display covariate imbalance. We also generate an outcome variable Y which is a function of the X covariate value as well as the treatment indicator t. This code also visualizes the data via a scatter plot of Y and X grouped by treatment status.

```
_____*
* Create data to analyze
*_____
                                    _____
*---> Set seed and initialize dataset
clear
set seed 123456
set obs 150
gen t = 1 if _n <= 50
replace t = 0 if t = =.
*---> generate a covariate where there are control observations with
*
     covariate values where no treatment observations exist
gen X = runiform(0, 10) if t == 0
replace X = runiform(5,10) if t == 1
*---> generate a dependent variable that is a function of X and t:
*
     A treatment effect of 1
*
     A nonlinear relation between Y and X for control observations.
gen Y = .5 + .5 \times X + 1 \times t + rnormal(0, 0.25)
replace Y = 3 + rnormal(0, 0.25) if X < 5
*---> plot the data to view what we've created
graph twoway (scatter Y X if t==0, leg(label(1 "control")) mcolor(black)) (scatter Y
X if t==1, leg(label(2 "treated")) mcolor(red) mfc(none))
*---> Graph covariate imbalance across treated and control
graph box X, over(t)
byhist X, by(t) bin(6) twl(color(red)) tw2(color(blue)) density
```



Using this code, we also plot a histogram of *X* separately by treatment status to depict covariate imbalance across the treated and control samples:



#### 3. Analysis Approaches

In this section we employ three common research designs to the simulated data to assess how each method weights observations in constructing an estimate of the treatment effect.

### 3.1. OLS Regression Only

By far, the most common approach accounting researchers to adjust for covariates is a multiple regression. In this approach, researchers estimate an ordinary least squares (OLS) regression over all available observations without missing data for the variables in their model. This approach treats all observations with an equal weight.

```
* Approach 1 - Regression only
*----> Estimate Regression
. regress Y t X
```

This produces the following output:

Source	SS	df	MS	Number of o	obs =	150
Model   Residual	146.342981 31.740546	2 147	73.1714905 .215922082	F(2, 147) $Prob > F$ $R-squared$	- = =	0.0000
Total	178.083527	149	1.19519146	Root MSE	=	.46467
Y	Coef.	Std. Err.	t	P> t  [959	& Conf.	Interval]
t   X   _cons	.8165832 .2897373 2.163907	.0866383 .0152053 .0909312	9.43 19.05 23.80	0.000 .645 0.000 .25 0.000 1.98	53657 59688 34206	.9878007 .3197865 2.343608

We see that the imbalance and nonlinearity result in a downward bias for the estimated coefficient on the treatment indicator t (0.817 coefficient vs 1.000 for an unbiased estimate) and on X (0.290 coefficient vs 0.500 for an unbiased estimate). This bias in both estimates derives from the assumption of a constant linear treatment effect across all observations in the sample, resulting in excess weight applied to control sample observations where X < 5 and the treatment effect is zero.

## 3.2. PSM and Regression

Accounting researchers' efforts to achieve covariate balance between treatment and control samples have emphasized propensity score matching (e.g., Rosenbaum and Rubin 1983; Armstrong, Jagolinzer, and Larcker 2010; Lawrence, Minutti-Meza, and Zhang 2011).<sup>1</sup> In this

<sup>&</sup>lt;sup>1</sup> Accounting researchers also employ alternative matching methods, including matching treated observations to control observations on a small subset of covariates using nearest neighbor matching within a pre-specified group such as industry (e.g., Kothari, Leone, and Wasley 2005), using coarsened exact matching to facilitate matching on several covariates (e.g., DeFond, Erkens, and Zhang 2017), or matching on a distance metric such as the generalized Mahalanobis distance (e.g., Diamond and Sekhon 2005).

approach, researchers estimate a propensity score model where the dependent variable is the treatment indicator and the independent variables are a set of covariates, compute propensity scores (p-scores) as the fitted values from this model, and then match treated observations to control observations with similar p-scores. With these subsamples identified, researchers estimate a regression over all matched observations. This approach results in unmatched observations receiving a zero weight and matched observations typically receiving a positive, integer weight. When matching is done without replacement, this weight will be one.

```
* Approach 2 - PSM - Caliper distance with replacement
*----> estimate Pscore
psmatch2 t X, norepl descending caliper(0.03) logit
*---> Examine '_weight'; weight variable created by psmatch2
tab _weight t
```

This code produces the following output:

First, it returns the estimated propensity score model. Fitted values from this model (p-scores) are used to construct a distance metric between observations (i.e., differences in p-scores). Matching typically proceeds by identifying a control sample observation for each treatment sample observation where the distance metric is the smallest and less than some threshold, often referred to as a caliper.

Logistic regression Log likelihood = -84.140984 t   Coef. Std. Err.				Number o LR chi2( Prob > o Pseudo F	of obs (1) chi2 R2	= = =	150 22.67 0.0000 0.1187
t   +	Coef.	Std. Err.	 Z	P> z	[95%	Conf.	Interval]
X   _cons	.3517719 -2.89988	.0827706 .5829605	4.25 -4.97	0.000 0.000	.1895 -4.042	445 462	.5139992 -1.757299

Next, this code describes the resulting weights \_weight from matching without replacement after sorting the data in descending order, and requiring a maximum difference between propensity scores of 0.03. Only 46 of the 50 treatment observations are matched via this process. This matched control sample is used as the counterfactual.

psmatch2:			
weight of matched		t	
controls	0	1	Total
1	+ 	46	+92 +
Total	46	46	92

Next we plot the data in these two samples.

```
*---> SCATTER PLOT
*plot scatter plot of matched
graph twoway (scatter Y X if t==0 & _weight<., leg(label(1 "P-score Matched"))
mcolor(black%30) xlabel(0(1)10) ylabel(2(1)7)) (scatter Y X if t==1 & _weight<.,
leg(label(2 "Treated")) mcolor(red) mfc(none))</pre>
```

Examining the scatter plot of Y and X of the matched observations, we can see the procedure discarded all control observations with values of X less than approximately 5, consistent with a valid match between treatment and control observations based on the single covariate.



We next estimate the regression after performing propensity score matching without replacement by only including those observations with a non-missing weight in the estimation.

```
*---> Estimate Regression
regress Y t X if _weight != .
```

This produces the following output:

Source	SS	df	MS	Number of obs	=	92 466,92
Model   Residual	80.6372937 7.68517487	2 89	40.3186468 .086350279	Prob > F R-squared	= = -	0.0000
Total	88.3224685	91	.970576577	Root MSE	=	.29385

Υ   +	Coef.	Std. Err.	t	P> t	[95% Conf.	Interval]
t	.9634928	.0613409	15.71	0.000	.8416098	1.085376
X	.5089889	.0200039	25.44	0.000	.4692415	.5487362
_cons	.4322711	.1499198	2.88	0.005	.1343837	.7301584

This analysis demonstrates that using propensity score matching to identify a matched sample eliminates bias in the regression estimate of the coefficients on t (0.963 vs 1.000 for an unbiased estimate) and on X (0.509 vs 0.500 for an unbiased estimate).

### 3.3. Entropy Balancing and Weighted Regression

While propensity score matching typically improves covariate balance, a more recently developed method of reweighting control sample observations, entropy balancing (Hainmueller 2012), virtually eliminates covariate imbalance with a simple, one-line Stata command. McMullin and Schonberger (2020) apply entropy balancing in the setting of estimating abnormal accruals to provide a detailed discussion of the advantages of entropy balancing in addressing non-linear relations with underlying covariates in order to eliminate (or at least reduce) bias in estimated treatment effects. Further, the authors describe several limitations of entropy balancing in empirical-archival research settings with features that are common in accounting research. We build on their discussion in the following illustration.

The first step in performing entropy balancing is to run the ebal command. This iterative algorithm creates a new variable \_webal, which when used to weight the control sample results in near perfect covariate balance. The option target(3) sets the balance conditions required to be met for the algorithm to cease adjusting the control sample weights, which in this case requires the first three moments of the X covariate distribution to be the same (within a default tolerance) across the treated and weighted control sample.

```
*-----*
* Approach 3 - Entropy Balancing and Weighted Regression
*---->run ebalance
ebalance t X, target(3)
```

This command produces the following output:

```
Data Setup

Treatment variable: t

Covariate adjustment: X (1st order). X (2nd order). X (3rd order).

Optimizing...

Iteration 1: Max Difference = 6186.94578

Iteration 2: Max Difference = 2276.21682

Iteration 3: Max Difference = 837.550557

Iteration 4: Max Difference = 308.317911

Iteration 5: Max Difference = 113.657737

Iteration 6: Max Difference = 42.0557396

Iteration 7: Max Difference = 15.8045807

Iteration 8: Max Difference = 6.60675262
```

Iteration 9: N	Max Differend	ce = 3.5086	0125			
Iteration 10:	Max Differer	nce = 1.949	48744			
Iteration 11:	Max Differer	nce = .7907	83647			
Iteration 12:	Max Differer	nce = .1962	87767			
Iteration 13:	Max Differer	nce = .0168	41045			
Iteration 14:	Max Differer	nce = .0001	39034			
maximum differ	cence smaller	than the	tolerance le	evel; conver	gence achiev	ved
				- · · · <b>,</b> · · · · ·	<b>J</b>	
Treated units:	: 50 tot	al of weig	hts: 50			
Control units:	: 100 tot	al of weight	hts: 50			
Before: withou	it weighting					
	jj					
		Treat		1	Control	
	mean	variance	skewness	mean	variance	skewness
	+			+		
х	7.25	2.243	.1802	5.14	8.323	.02468
'						
After: webal	L as the weig	hting vari	able			
		,, , ,				
		Treat		1	Control	
	mean	variance	skewness	mean	variance	skewness
1				+		
x	7.25	2,243	.1802	7,25	2,243	.1802
	,.13		1001	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	2.210	1002

The first panel provides summary statistics that allow us to assess pre-adjustment covariate balance. The second panel shows the summary statistics after weighting the control sample using weights identified by the entropy balancing algorithm. While covariate balance was achieved, we next examine the weights created to achieve the balance.

```
*---> Analysis examining entropy balancing weights
. tabstat _webal, statistics(mean, max, N) by(t)
. gsort - t _webal
. list _webal if inrange(_n, 1, 10) & t == 0 | inrange(_n, _N-10+1, _N)
```

This produces the following output:

From this output we can see that the entropy balancing algorithm retains all 100 control and all 50 treatment observations. This output also shows that the treatment sample observations retain a

weight of one and that continuous weights assigned to control sample observations range from nearly zero (1.27e-18) to 1.56.

-	++
	webal
141.	1.3520483
142.	1.4549744
143.	1.4727891
144.	1.4858081
145.	1.5137473
146.	1.5182191
147.	1.5204759
148.	1.5512371
149.	1.5558077
150.	1.5623861
-	++

This list of the top 10 weights shows that no single observation was assigned an excessively high weight.

Given that we only have one covariate in this simulated example, we can compare the weight assigned to each observation and its value of X to understand how the weights are iteratively adjusted. To do this we use the following code create a scatter plot of \_webal and X after each of the 14 iterations required to achieve covariate balance within the default tolerance of 0.015.

```
*---> Plot covariate X and entropy balance created variable '_webal'
graph twoway (scatter _webal X if t==0, leg(label(1 "Entropy Balanced"))
mcolor(blue) mfc(none) mlw(vthin) ysc(r(0 1.6)) ylabel(0(.2)1.6, angle(0) grid) )
(scatter _webal X if t==1, leg(label(2 "Treated")) mcolor(red) mfc(none))
```



These plots show the entropy balancing weights after N iterations.<sup>2</sup>

<sup>&</sup>lt;sup>2</sup> We achieve this series of plots by stopping ebalance from iterating by increasing the tolerance. (E.g., setting tolerance to 7000 (i.e., adding the option tol(7000)) results in no adjustment, setting it to 5000 results in 2 iterations of adjusting the weights, 2000 results in 3 iterations, etc.) This tolerance corresponds to the Max Difference parameter reported with each iteration. See the accompanying Stata do file for the remaining tolerances we specify to stop the algorithm after each of the N iterations.

We direct the reader interested in the theoretical underpinning of this entropy optimization algorithm to Kapur and Kesavan (1992) or Mattos and Veiga (2004). Hainmuller (2012) provides these references for more detail on the iterative algorithm he employs in this technique.

Examining these plots, we see the iterative algorithm decreased weights assigned to observations with an *X* value less than 4 to be nearly zero. The remainder of the weights were assigned following a continuous function that spans the range of *X* values in the treatment sample.

To visualize the weights assigned by entropy balancing after the final iteration, we construct the scatter plot of the covariates *Y* and *X* for all observations in the sample, weighted by \_webal.

```
*---> SCATTER PLOT
* Ensure Stata produces consistent sized dots based on weights across both groups
* See https://www.stata.com/statalist/archive/2008-08/msg00987.html
expand 2
replace X = . if _n>(_N/2)
recode t (1=0) (0=1) if X==.
*plot scatter plot of Y and X based on entropy balancing weights (_webal)
graph twoway (scatter Y X if t==0 [aw=_webal], leg(label(1 "Entropy Balanced"))
mcolor(blue) mfc(none) mlw(vthin) msize(*.25)) (scatter Y X if t==1 [aw=_webal],
leg(label(2 "Treated")) mcolor(red) mfc(none) msize(*.25))
```

This generates the following plot:



We see that all observations are retained after entropy balancing. This approach also downweighted control observations where there was no overlap with treatment sample (i.e., X values below 5), assigning a weight of effectively zero to observations with X values below 4.

The estimation of the regression after executing the entropy balancing procedure is done using the pweight option and the \_webal variable.

```
*---> Estimate Regression
. regress Y t X [pweight=_webal]
```

This produces the following output:

(sum of wgt is	1.0000e+02	2)				
Linear regress	ion			Number of F(2, 147) Prob > F R-squared Root MSE	obs = = = = =	150 456.37 0.0000 0.9099 .28543
 У	Coef.	Robust Std. Err.	t	P> t	[95% Conf.	Interval]
+ t   X   _cons	.9686133 .5085347 .4256848	.0574659 .0188487 .1463198	16.86 26.98 2.91	0.000 0.000 0.004	.8550472 .4712853 .1365227	1.082179 .5457841 .7148468

Similar to the PSM approach, this demonstrates that entropy balancing is effective in eliminating bias in the estimation of the coefficients on t (0.967 vs. 1.000 for an unbiased estimate) and on X (0.509 vs. 0.500 for an unbiased estimate). While these estimates retain all observations in the dataset, the impact of those control observations that are not similar to the treated observations is effectively zero. This implies that the counterfactual is constructed by weighting the observations in the control samples using the entropy balance weights and that this counterfactual is similar to the treated sample.

#### References

- Armstrong, C., A. Jagolinzer, and D. Larcker. 2010. "Chief Executive Officer Equity Incentives and Accounting Irregularities." *Journal of Accounting Research* 48 (2): 225-271.
- DeFond, M., D. H. Erkens, and J. Zhang. 2017. "Do Client Characteristics Really Drive the Big N Audit Quality Effect? New Evidence from Propensity Score Matching." *Management Science* 63 (11): 3531–3997.
- Diamond, A., and J. S. Sekhon. 2005. "Genetic Matching for Estimating Causal Effects: A General Multivariate Matching Method for Achieving Balance in Observational Studies." *Review of Economics and Statistics*, no. 0.

- Hainmueller, J. 2012. "Entropy Balancing for Causal Effects: A Multivariate Reweighting Method to Produce Balanced Samples in Observational Studies." *Political Analysis* 20 (1): 25–46.
- Kapur, J. N., and H. K. Kesavan. 1992. Entropy Optimization Principles with Applications. Springer.
- Kothari, S. P., A. J. Leone, and C. E. Wasley. 2005. "Performance Matched Discretionary Accrual Measures." *Journal of Accounting & Economics* 39 (1): 163–97.
- Lawrence, A., M. Minutti-Meza, and P. Zhang. 2011. "Can Big 4 versus Non-Big 4 Differences in Audit-Quality Proxies Be Attributed to Client Characteristics?" *The Accounting Review* 86 (1): 259–86.
- Mattos, R., and A. Veiga. 2004. "Entropy Optimization: Computer Implementation of the Maxent and Minexent Principles." Working Paper. Universidade Federal de Juiz de Fora, Brazil.
- McMullin, J. L., and B. Schonberger. 2020. "Entropy-Balanced Accruals." *Review of Accounting Studies* 25 (1): 84–119.
- Morgan, S. L., and C. Winship. 2015. *Counterfactuals and Causal Inference*. Cambridge University Press.
- Rosenbaum, P. R., and D. B. Rubin. 1983. "The Central Role of the Propensity Score in Observational Studies for Causal Effects." *Biometrika* 70 (1): 41–55.